

AN UPDATED STATUS OF ALANINE RACEMASE INHIBITORS: A REVIEW

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ABSTRACT: The predominant function of the enzyme alanine racemase in the formation of mycobacterium cell walls has brought it widespread recognition. Alanine racemase produces D-alanine, a peptidoglycan precursor that is absolutely necessary for the proper development and maintenance of the cell wall. There is great promise in the lipid-rich mycobacterium cell wall as a therapeutic target for the development of novel drugs, as it is common among prokaryotes. Inhibiting Alanine racemase should be the top priority when dealing with different pathogenic infections because of its critical function in mycobacterium - cell wall formation. Renewed attempts to find novel and improved inhibitors with superior therapeutic indices were driven by concerns about cellular toxicity, interference with metabolic processes, and lack of selectivity produced by numerous recognized inhibitors. For the benefit of humanity, this study summarizes the current state of reported Alanine racemase inhibitors based on fragments of evidence in the literature. The goal is to facilitate the rationalization of the drug discovery process as a whole by facilitating the exploration, design, and identification of more precise inhibitors.

Keywords: In-situgel, Acyclovir, Anti-viral, HPMCE50 LV, PluronicF-127

INTRODUCTION: Microorganisms are defined as infectious agents of microscopic size, including bacteria, fungi, protozoan and viruses, responsible for causing various types of infections. To deal with infectious agents, there is an urgent need for an antimicrobial agent that antagonizes the action of infection-causing microbe¹. The

discovery of antimicrobial drugs conferred huge benefits on human health and changed the fate of mankind dramatically. Penicillin was the first antibiotic discovered by Alexander Fleming, which proved to be a boon in curing infectious diseases. As a result, antibiotics were regarded as wonder drugs and generally used to manage infections caused by pathogens. However, a

overconsumption, which is a key issue of concern for the researchers

².

Now the greatest challenge of the twenty-first century is the development of drug resistance responsible for causing immense human suffering. The resistance problem urges iterated efforts to strive for antibacterial agents efficacious against pathogenic bacteria rebellious to commercial antibiotics

^{3,4}. This highlights the immediate call for upgraded antibacterial agents with advanced mechanisms for clinical application^{5,6}.

Potential Target Sites for the Search of Futuristic Antimicrobials:

The treatment of infectious disease becomes knotty as microbial resistance shoots up at odds with antimicrobial agents. Drugs that destroy microbes prevent their proliferation, and pathogenic actions have dissimilar structures, inconsistent affinity towards the target site, and disparate spectrum of activity with various mechanisms of action. The

advancement in bacterial genomics has greatly altered the antimicrobial therapeutic environment, and many potential targets stand by^{7,8}.

modern drugs acting via advanced mechanisms or against the latest target sites from natural sources⁹. The

probable targets for searching for new antimicrobial compounds may be focused on the following mechanisms⁸, which are depicted in Fig. 1 as follows.

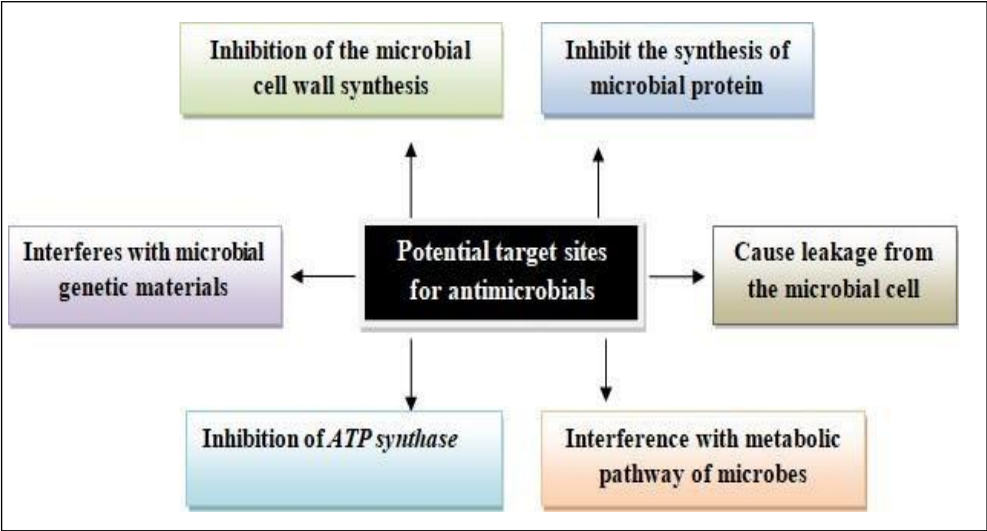


FIG.1:POTENTIAL TARGETSITESFORANTIMICROBIALAGENTS

Inhibition of Microbial Cell Wall Synthesis: Bacterial cells are surrounded by a cell wall made of a peptidoglycan network constitutes an essential component of the cell wall, serves as a perfect site for drug designs since corresponding biosynthetic

process are lacking in mammalian hosts. Blockage of bacterial cell walls synthesis is of paramount importance for the action of antimicrobials. Their probable target sites^{10-13,8} in cell wall synthesis are summarized in Fig. 2 maybe.

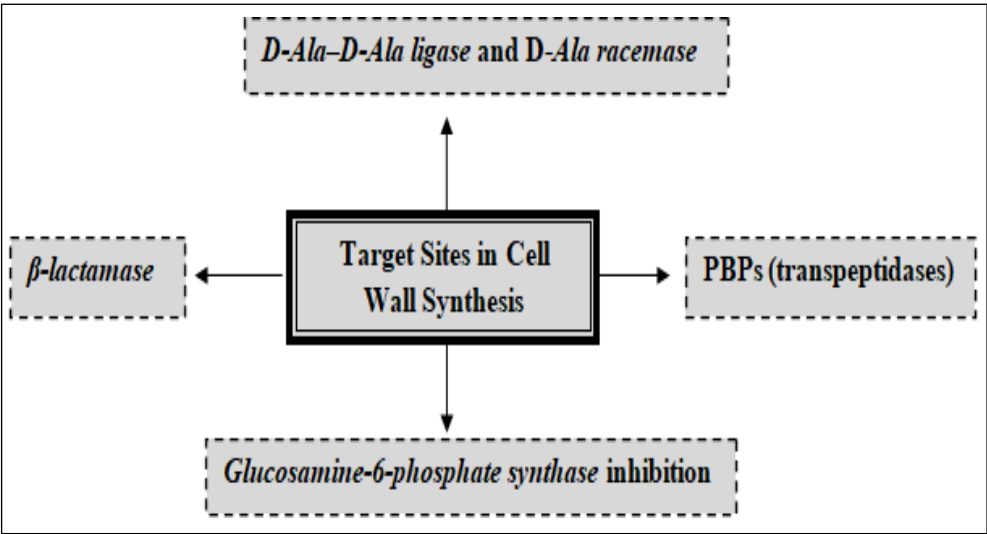
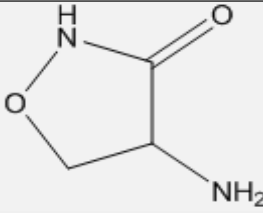
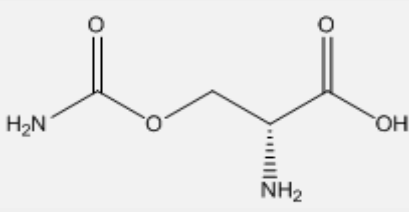
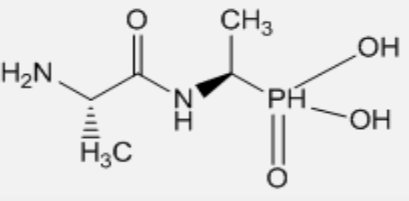
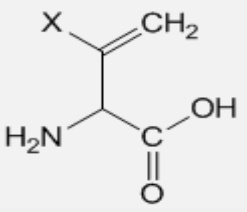


FIG.2:TARGETSITESINCELLWALLSYNTHESIS

AlanineracemasePromisingTargetforAntimicrobial agents: *Alanine racemase*(Alr, EC5.1.1.1)isapyridoxal-5-phosphate(PLP)dependent homodimeric enzyme that brings aboutreversibleracemizationofL-alanineandD-alanine.Thisbacterialenzymeexecutepredominating role in cell wall synthesis of bacteria^{14,15}byproviding D-alanine(D-ala) whichserveasakeymoleculeforthebiosynthesisofpeptidoglycan network of mycobacterial cell wall;hence its inhibition has been reported to be fatal topathogen viability in the deprivation of D-alaninesupplementation ^{16,17}. The lipid-rich mycobacterialcell wall is common amidst prokaryotes, making*Alanine racemase*a putative target for the designand development of pharmacologically active drug¹⁸⁻²¹.D-alanineprovidedby*Alanineracemase*is

vital for maintaining cell wall growth and integrity.D-alanineactsasapivotalprecursorforpeptidoglycan biosynthesis in bacterialcell wallsvia D-ala-D-ala formed by the enzyme D-ala-D-alaligase²².Thismanifestshowtheinhibitionof*alanineracemase*isimportunate.ThispaperprovidesanoverviewoftheupdatedstatusofreportedAlanineracemaseinhibitorsbasedonshredsofliterature.The productsderivedfromnaturalssourceshavebeenrecognizedtoplayasignificant role by being the lead molecules to beselectedaspotentialcandidatesfordrugdevelopment. Various researchers have synthesizedderivativesofdifferentscaffoldsandevaluatedthem for *Alanine racemase*properties, summarizedin **Table 1**.

TABLE1:REPORTEDINHIBITORSOFFENZYME ALANINERACEMASE

Sr.no.	ReportedInhibitors	ResearchFindings
		Sources: <i>Streptomyces garyphalus</i> or <i>S. orchidaceus</i> . Dissociation
		
		
		

1.D-cycloserine(DCS)

2.O-carbamyl-D-serine

3.Alafosfalin

followed by subsequent rearrangement of DCS with substituted oxime untriddealanine racemase reactivation in cellular pool. DCS, earlier proved to be an effective competitive inhibitor of enzyme, unfit for *S. aureus* Alr due to the absence of conformation essential for the molecule to bind with substrate region²³⁻²⁵.

Enzyme kinetics-

 $K_m = 4.6 \times 10^{-4} \text{ M (D-alanine)}$ $K_m = 9.7 \times 10^{-4} \text{ M (L-alanine)}$ Sources: *Streptococcus faecalis*

Determination of primary site of action of O-carbamyl-D-serine on Alr on the basis of UDP-NAC muramyl-L-ala-D-glu-L-lys accumulation and in the absence of D-ala-O-carbamyl-D-serine^{26, 27}

Enzyme kinetics- $K_m = 4.8 \times 10^{-4} \text{ M (D-alanine)}$, $K_m = 6.8 \times 10^{-3} \text{ M (L-alanine)}$

Sources: Synthetic L-alanine analog

Contains two parts-Ala R inhibitor of fosfalin and carrier alanine moiety

Based upon alafosfalin information of external aldimine with PLP cofactor, phosphonate group rendered catalytic residues inaccessible for catalysis.

Variable activity against gram positive and negative bacterial strains.

Phosphonodipeptide with antibacterial properties²⁷⁻²⁹

4.

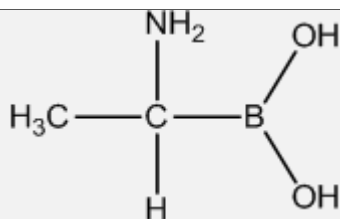
Sources: synthesized from N-(benzyloxycarbonyl)-vinylglycine methyl ester which in turn obtained from methionine. Irreversible inhibitor of Alr obtained from *E. coli*.^{27,30}

D-chlorovinylglycine:

MIC value- 32 $\mu\text{g/mL}$ (*S. aureus*) 64 $\mu\text{g/mL}$ (*S. faecalis*)

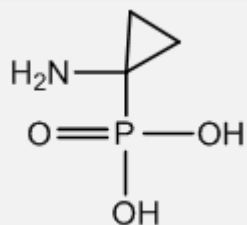
X=Cl, F Halovinylglycine

5.



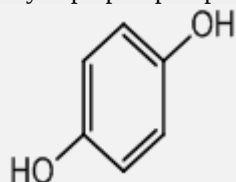
(1-aminoethyl)boronic acid

6.



1-aminocyclopropanephosphonate

7.



Hydroquinone

Structural analog of alanine

Showed time-

dependent inhibitory activity towards Alr isolated from *B. stearothermophilus*

Competitive inhibitor of D-alanine at D-alanine binding sites^{27,31}

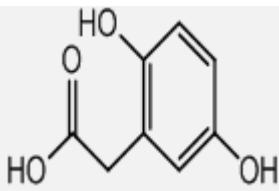
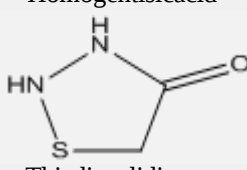
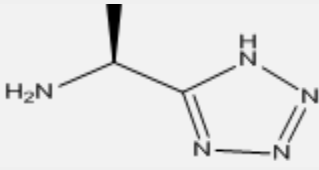
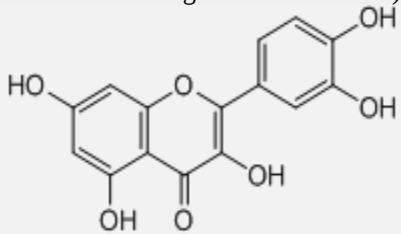
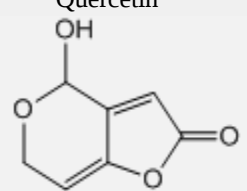
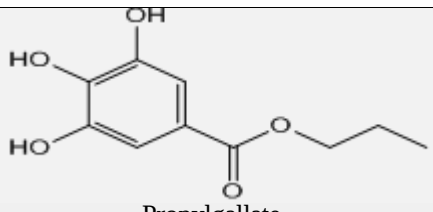
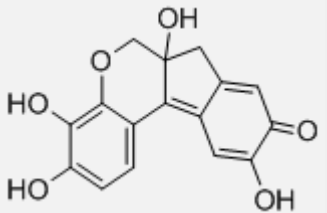
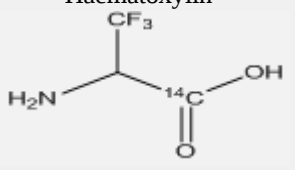
Slow but potent inhibitor of Alr obtained from *B. stearothermophilus* strain.

Second order rate constant: $< 150 \text{ M}^{-1} \text{ s}^{-1}$ $^1K_m/K_i$ ratio: 2000^{27,32}

Sources: Blueberry, pears, broccoli, onions, tea, coffee, beer, red wine, wheat and cereal etc.

Alr-2 inhibitors found to be active against *Aeromonas hydrophila*IC₅₀ value= 11.39 μM MIC value= 25 $\mu\text{g/mL}$

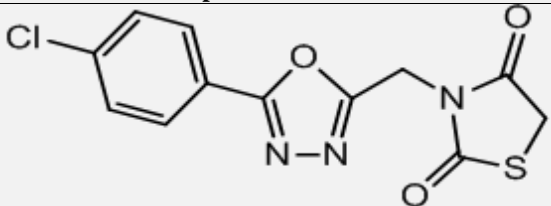
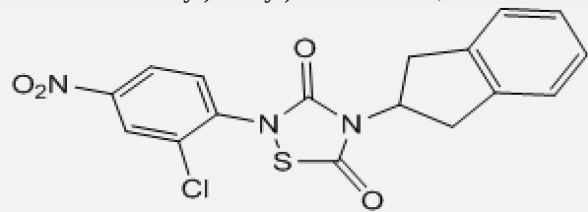
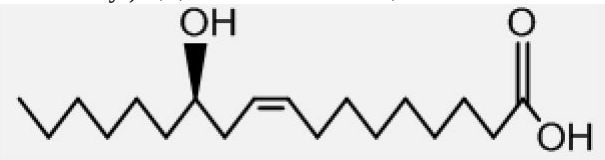
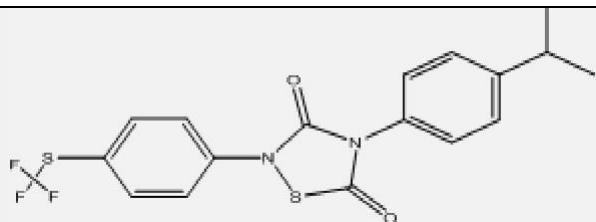
Potent inhibitor of Alr isolated from *Streptococcus iniae* HNM-1^{18,33}

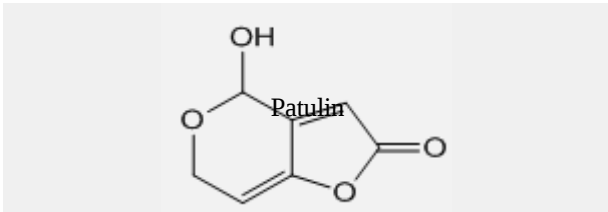
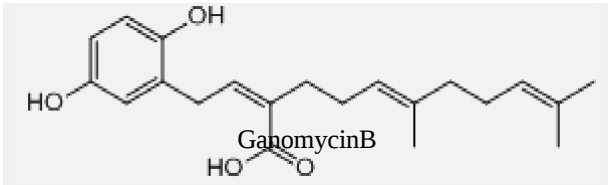
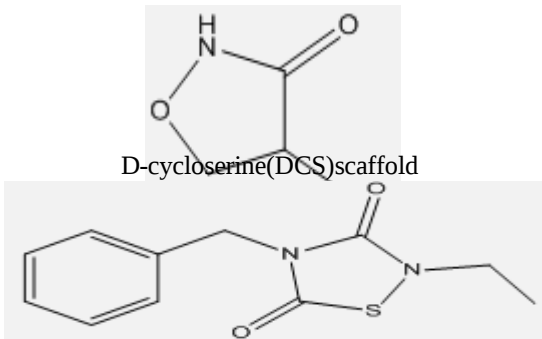
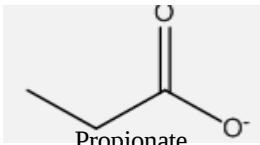
8.		Sources: Aebutusunedo (strawberry-tree), honey, <i>Xanthomonas campestris</i> spv. Phaseoll, Yarrownia lipolytica Alr-2 inhibitors found to be active against <i>Aeromonas hydrophila</i> IC ₅₀ value = 0.2 μM MIC value = 1.73 μg/mL Potent inhibitor of Alr isolated from <i>Streptococcus iniae</i> HNM-1 ^{18,33,34}
9.		Inhibitory activity against <i>M. tuberculosis</i> and <i>M. smegmatis</i> Alr with IC ₅₀ value ranging from <0.03 to 28 μM and 23 to >150 μM respectively ³⁵ .
10.		C-terminal 1-aminoethyltetrazole containing di and oligopeptides synthesized by solid phase peptide coupling technique were identified as novel and potential <i>alanine racemase</i> inhibitors ²⁹ .
11.		Sources: Apples, berries, broccoli, grapes, citrus fruits, cherries, green tea, coffee, red wine, onions and capers etc. M.W = 302.24 IC ₅₀ value = 0.33 μM Docking score = -102.489 kcal/mol Potent inhibitor of AlaR Effective drug in the treatment of pulmonary tuberculosis ^{18,36}
12.		Sources: Apple, peach, grapes, fruit juices, several species of <i>Aspergillus</i> , <i>Penicillium</i> and <i>Byssoschlamys</i> ^{18,37-39} M.W = 154.12 IC ₅₀ value = 14.7 (0.27) μM MIC value = 20 (0.89) μg/mL
13.		M.W = 212.2 IC ₅₀ value = 8.6 (0.5) μM ¹⁸
14.		Sources: Heartwood of the logwood tree <i>Haematoxylum campechianum</i> ⁴⁰ M.W = 302.29 IC ₅₀ value = 15.6 (0.23) μM
15.		Exhibited time-dependent inhibition of <i>AlaR</i> obtained from <i>S. typhimurium</i> and <i>B. stearothermophilus</i> ²⁷ K _i values = 65 ± 10 and >100 mM K _{ina} α = 0.08 ± 0.02 and >2.0 min ⁻¹

In drug discovery, computer-aided drug design (CADD) offers effective and reliable methodologies for lead optimization, virtual screening, and designing new drug candidates. Molecular docking is a computational drug design approach that provides insights into molecular recognition⁴¹. In an attempt to conduct wet laboratory experiments smoothly and effectively, this method is useful for predicting the

compound architecture, preferred orientation and conformation (binding pose), interaction, and binding geometry of small ligands into the catalytic pocket of biomolecular targets based on docking score and function⁴². Based on literature evidence, molecular docking studies of some reported *Alanine racemase* inhibitors have been presented in **Table 2**.

TABLE 2: REPORTED MOLECULAR DOCKING-BASED STUDIES OF ALANINE RACEMASE INHIBITORS

Sr.no.	Reported Inhibitors	Research Findings
1	 3-((5-(4-chlorophenyl)-1,3,4-oxadiazol-2-yl)methyl)thiazolidine-2,4-dione	PDB code: 1XFC Docking software- Schrodinger ⁴³ IC ₅₀ value =13.1 μM Number of H- bonds-6 Salt bridge interaction-Arg228
2	 2-(2-chloro-4-nitrophenyl)-4-(2,3-dihydro-1H-inden-2-yl)-1,2,4-thiadiazolidine-3,5-dione	PDB code: 1XFC IC ₅₀ value=0.05 μM Number of H-bonds-3 π-π stacking-His172, Tyr175 ⁴³
3	 Ricinoleic acid	PDB code: 3E5P Docking software- AutoDock Docking score: -7.81, Number of H-bonds- 5 Amino acid residues – Ser207, Val225, Tyr356, Tyr267. Bioavailability score: 0.56 with high GI absorption ⁴⁴
4		PDB code: 1XFC, Resolution: 1.9 Å Docking software- Schrodinger Induced fit docking- Desmond Induced fit docking results revealed that Lys42, Tyr46, Tyr175 and Tyr364 residues were responsible for the stabilization of inhibitor-protein complexes ⁴⁵

5	4-[4-(propan-2-yl)phenyl]-2-{4-[(trifluoromethyl)38.88kcal/mol sulfanyl]phenyl}-1,2,4-thiadiazolidine-3,5-dione	Binding energy value = - IC ₅₀ value = 0.17 µM PDB code: 2rjh.1.A Docking software: AutoDock Compound possessed inhibitory activity against <i>Aeromonas hydrophilla</i> ¹⁸ IC ₅₀ value = 0.62 µM against Caco-2 cells Also exhibits strong cytotoxic effects and reduces the viability of HeLa cells up to 99% at 6.25 µg/ml.
6		PDB code: 2RJG Docking software: AutoDock 4 Predicted Xscore Ki = 0.15 µM Compound forms H-bonds with residues Arg280, Tyr274 and prosthetic group PLP. Compound had narrow access to the active site ⁴⁶
7		PDB code: 1XFC-A (Mtb-Alr) Software: MODELLER (Homology modelling) Docking software: AutoDock Vina ¹⁵ Molecular Volume: 205.03
8	 4-benzyl-2-ethyl-1,2,4-thiadiazolidine-3,5-dione	PDB code: 1SFT Program: LigBuilder (to generate pharmacophore model) Non-covalent inhibitor fit only two features of the dynamic pharmacophore model with no excluded volumes, made the compound insufficient in order to be oriented onto the model with excluded volumes ⁴⁷
		

CONCLUSION: The imperative role of alanine racemase in mycobacterium cell wall synthesis

implies that its inhibition is of utmost priority in dealing with various pathogenic infections. This paper provides an overview of structural information on various reported inhibitors of alanine racemase based on convincing shreds of evidence from literature so that more precise inhibitors could be explored, designed and identified to rationalize the overall drug discovery process which will be true serendipity for humankind.

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